

CHAPTER 1

# The onset of fever: new insights into its mechanism

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**Abstract:** The classical view of fever production is that it is modulated in the ventromedial preoptic area (VMPO) in response to signaling by pyrogenic cytokines elaborated in the periphery by mononuclear phagocytes and the consequent induction of cyclooxygenase (COX)-2-dependent prostaglandin (PG)E<sub>2</sub> in the VMPO. This mechanism has, however, been questioned, in particular because the appearance of circulating cytokines lags the onset of the febrile response to intravenously (iv) injected bacterial endotoxic lipopolysaccharide (LPS), an exogenous pyrogen. Moreover, COX-2, in this case, is itself an inducible enzyme, the de novo synthesis of which similarly lags significantly the onset of fever. Issues also exist regarding the accessibility of the POA to blood-borne cytokines. New data adduced over the past 10 years indicate that the peripheral febrigenic message is conveyed to the VMPO via a neural rather than a humoral route, specifically by the vagus to the nucleus tractus solitarius (NST), and that the peripheral trigger is PGE<sub>2</sub>, not cytokines; vagal afferents express PGE<sub>2</sub> receptors (EP<sub>3</sub>). Thus, the initiation of the febrile responses to both iv and intraperitoneal (ip) LPS is temporally correlated with the appearance of LPS in the liver's Kupffer cells (Kc), its arrival immediately activating the complement (C) cascade and the consequent production of the anaphylatoxin C5a; the latter is the direct stimulus for PGE<sub>2</sub> production, catalyzed non-differentially by constitutive COX-1 and -2. From the NST, the signal proceeds to the VMPO via the ventral noradrenergic bundle, causing the intrapreoptic release of norepinephrine (NE) which then evokes two distinct core temperature ( $T_c$ ) rises, viz., one  $\alpha_1$ -adrenoceptor (AR)-mediated, rapid in onset, and PGE<sub>2</sub>-independent, and the other  $\alpha_2$ -AR-mediated, delayed, and COX-2/PGE<sub>2</sub>-dependent, i.e., the prototypic febrile pattern induced by iv LPS. The release of NE is itself modulated by nitric oxide contemporaneously released in the VMPO.

**Keywords:** pyrogenic agents; preoptic-anterior hypothalamus; vagal afferents; Kupffer cells; prostaglandin E<sub>2</sub>; complement 5a; norepinephrine; nitric oxide

## Introduction

The conventional view of the mechanism by which infectious fevers are produced holds that infectious noxa (e.g., Gram-negative bacteria and/or their products [bacterial endotoxic lipopolysaccharides,

LPS]) that invade the body activate mononuclear phagocytes that then produce and release pyrogenic cytokines. These, in turn, are transported by the bloodstream to the ventromedial preoptic area (VMPO) of the anterior hypothalamus, the “fever-producing center”, where they act (for reviews, see Saper, 1998; Roth and De Souza, 2001; Dunn, 2002; Dinarello, 2004). It is generally agreed that evidence of their central action is the depression of

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warm-sensitive neurons in this region, leading, in accordance with Hammel's classical model (1965), to the conservation of heat and, hence, to a rise in body core temperature ( $T_c$ ) (Boulant, 2000). It is, however, uncertain how cytokines, as hydrophilic peptides, could penetrate the brain and, indeed, whether they or other, secondarily elaborated factors mediate the febrile response. To wit, it is generally believed that, rather than acting directly, the cytokines induce the local generation and release of prostaglandin (PG) $E_2$ , a lipid mediator that is clearly thermogenic when injected centrally (for reviews, see Blatteis, 1997; Ivanov and Romanovsky, 2004). Its production in this instance has been demonstrated to be dependent on the activation of two enzymes, cyclooxygenase (COX)-2 and microsomal PGE synthase (mPGES)-1, which catalyze its conversion from arachidonic acid (AA) present in the membranes of cells (Ivanov et al., 2002).

Questions have arisen over the past 10 years, however, about the validity of this concept. They are based, in particular, on findings that low-to-moderate doses of LPS injected intravenously (iv) elicit in many species, e.g., guinea pigs (Sehic et al., 1996a, b), significant elevations of  $T_c$  and VMPO PGE $_2$  levels within 10 min after administration, whereas the first cytokine to appear in the blood, tumor necrosis factor (TNF) $\alpha$ , is not detectable until, at the earliest, 30 min after injection (Jansky et al., 1995). This would make it unlikely, therefore, that TNF $\alpha$  or other pyrogenic cytokines released subsequently could play a major role in the quick induction of these responses. This is not completely surprising, however, since cytokines are not generally expressed constitutively in mononuclear phagocytes, but are induced de novo in response to these cells' activation by LPS, a synthetic process that requires some time (Conti et al., 2004). By the same token, it has been demonstrated that both the COX-2 and mPGES-1 that catalyze the production of preoptic PGE $_2$  in response to peripheral LPS are also not constitutive, but rather are upregulated after a ca. 60 plus-min delay (Inoue et al., 2002). Hence, it would also seem improbable that PGE $_2$  elaborated by this mechanism could mediate the initiation of the febrile response. A different mechanism must therefore be operating to account for the prompt onset of fever after iv LPS.

### Pyrogen activation of the brain: the role of the vagus

The first part of the old concept that can probably be rejected is the notion that the cytokines are transported to the brain by the circulation for their action. If they are not detectable in the bloodstream before or coincidentally with the beginning of fever, their signaling of the brain is most likely not related to their physical arrival at this site. Indeed, if they had a role at all in initiating fever, the only pathway that would allow the rapid and direct transmission of their message from the periphery to the brain would be neural. Moreover, since the liver contains the body's largest population of mononuclear phagocytes (Kupffer cells, Kc), it is the primary clearinghouse of circulating LPS and therefore also the principal source of LPS-induced cytokines; the pyrogenic signal should therefore originate there. Indeed, the essential role of Kc in fever production was recently verified (Feleder et al., 2003; Li and Blatteis, 2004; Li et al., 2004). Furthermore, a number of studies have now established that the subdiaphragmatic section of both vagal trunks, and its hepatic branches in particular, significantly attenuates the febrile response to LPS (Watkins et al., 1995; Sehic and Blatteis, 1996; Romanovsky et al., 1997a, b; Fleshner et al., 1998; Gaykema et al., 1998; Simons et al., 1998; Wiczorek et al., 2005), albeit that it is still controversial whether this effect is conditional on the route and dose of LPS administered (Goldbach et al., 1997; Hansen et al., 2001). Biotinylated IL-1-receptor antagonist has also been shown to bind to glomus cells in hepatic vagal paraganglia (Goehler et al., 1997), suggesting that this could be the mechanism by which IL-1 $\beta$  activates the vagal afferents. In support, the intraportal vein administration of IL-1 $\beta$  increases the discharge rate of vagal afferents (Nijima, 1996) and the expression of *c-fos* in the nucleus of the solitary tract (NTS, the primary projection area of the vagal nerves) is enhanced after iv and intraperitoneal (ip) IL-1 $\beta$  and iv LPS (Wan et al., 1994). Subdiaphragmatic vagotomy abrogates this effect and electrolytic lesions of the NTS attenuate the febrile response to ip LPS (Wan et al., 1994). However, since Kc do not express cytokines constitutively, they cannot be the factors that rapidly

stimulate these terminals after LPS administration. Consequently, we should look for another factor that is quickly liberated in response to circulating LPS and capable of binding to vagal afferents to mediate the febrile response.

### **PGE<sub>2</sub>, not cytokines, is the peripheral fever trigger**

Such a factor could be PGE<sub>2</sub>. It is produced by Kc activated by LPS and its level rises in venous blood and, to a lesser extent, in arterial blood very quickly after the peripheral administration of both exogenous (e.g., LPS) and endogenous (e.g., IL-1 $\beta$ ) pyrogens (Skarnes et al., 1981; Rotondo et al., 1988; Perlik et al., 2005; Li et al., 2006). This raises the possibility that, as was conceived earlier in regard to the cytokines, the PGE<sub>2</sub> that acts in the VMPO could enter it from the blood: being lipophilic, it could either cross the blood-brain barrier (BBB) or diffuse to this site through “leaky” ports in the barrier, e.g., the organum vasculosum laminae terminalis (Blatteis and Sehic, 1997). It is, however, controversial whether PGE<sub>2</sub> can actually pass from the blood into the brain and, especially, whether PGE<sub>2</sub> entering the brain in this manner can raise  $T_c$  (Morimoto et al., 1992; Sehic et al., 1996a, b; Abul et al., 1997; Romanovsky et al., 1999). Indeed, although the rise of  $T_c$  and preoptic PGE<sub>2</sub> levels induced by iv LPS are both abrogated by peripherally injected antipyretics, findings that they are also prevented by the intraVMPO microinjection of COX-2 inhibitors would indicate that it is more likely that the applicable PGE<sub>2</sub> is generated inside rather than outside the BBB (Steiner et al., 2001). Hence, again, it would seem more probable that its message should be rapidly conveyed from the liver to the brain neurally rather than humorally. In support, abundant PGE<sub>2</sub> receptors of the EP<sub>3</sub> subtype occur in nodose ganglion vagal sensory neurons receiving information from the abdominal compartment (Ek et al., 1998).

There is a problem, however, with the notion that PGE<sub>2</sub> produced by Kc in quick response to LPS is the candidate mediator of the febrile response, because LPS is actually a weak trigger of AA release, the activation of group IV cytosolic phospholipase A<sub>2</sub> (cPLA<sub>2</sub>) by LPS being very slow

(Ambs et al., 1995). Moreover, as indicated earlier, the increased biosynthesis of PGE<sub>2</sub> induced by LPS is selectively catalyzed by inducible COX-2 and mPGES-1, and, in conscious rats at least, their mRNAs are not detectable in liver until 30 min after iv LPS (Ivanov et al., 2002). The delay imposed by this synthetic process consequently implies that the prompt elevation of plasma PGE<sub>2</sub> levels observed in response to iv LPS cannot be accounted for by its COX-2/mPGES-1-mediated production in Kc. A different factor must therefore be involved.

This mediator is the anaphylatoxic complement (C) component C5a (for review, see Blatteis et al., 2004a, b). The C cascade is activated on contact by LPS via the alternative pathway, resulting in the very quick production of all its components, including C5a. Kc express its cognate receptor, C5aR<sub>1</sub> (Schieferdecker et al., 1997, 2001; Schlaf et al., 2003). The production of PGE<sub>2</sub> by Kc is initiated within minutes after the addition of C5a, both *in vitro* and *in vivo*; C depletion inhibits this release (Schieferdecker et al., 2001). PGE<sub>2</sub>, under these conditions, could be generated by the hydrolysis of membrane-associated phosphoinositide (PI, which has a high arachidonoyl chain content) by PI-specific phospholipase C (PI-PLC); indeed, AA liberation by PI-PLC is 10-fold more rapid (within seconds) than that mediated by cPLA<sub>2</sub> (Rhur, 1994). Moreover, PI-PLC is activated by C, but not by LPS or IL-1 $\beta$ , and the subsequent conversion of this AA to PGE<sub>2</sub> is unselectively catalyzed by COX-1 and/or COX-2 (Schütze and Krönki, 1994), which are both constitutive in Kc. Hence, the initial peripheral fever trigger could indeed be PGE<sub>2</sub> released by Kc stimulated by LPS-activated C5a and binding to EP<sub>3</sub> receptors on vagal afferents.

This hypothesis was recently substantiated in a series of studies (for review, see Blatteis et al., 2004a, b; Perlik et al., 2005). Thus, the intraportal vein injection into anesthetized guinea pigs of LPS or cobra venom factor (CVF, another immediate activator of the C cascade that quickly elevates the levels of all the C components, but, unlike LPS, that eventually depletes the C substrate, thereby ultimately causing hypocomplementemia) induced similar increases of PGE<sub>2</sub> in the inferior vena cava (ivc, near its confluence with the hepatic veins) within the first 5 min after treatment. The rises in PGE<sub>2</sub> due to

LPS → Liver → C5a → Kupffer cells → PGE<sub>2</sub>

Fig. 1. Depiction of the sequence of steps that occur in the periphery following the iv or ip bolus injection of a small-to-moderate dose of LPS into conscious guinea pigs, culminating in the rapid production and release by Kc of PGE<sub>2</sub>, the endogenous trigger of the febrile response. Pyrogenic cytokines do not play a role in this phase of fever onset.

CVF returned to control levels within 15 min, whereas those due to LPS increased further 30–45-min later, then stabilized. LPS given to the same animals 3 h after CVF (when the pool of C5a was exhausted) also elevated ivc PGE<sub>2</sub>, but after a 30–45-min delay. CVF *per se* did not affect the basal level of ivc PGE<sub>2</sub>. It also did not alter basal ivc TNF $\alpha$ , IL-1 $\beta$ , and IL-6 levels nor their responses to LPS; in the latter case, their rise lagged significantly that of PGE<sub>2</sub>. These results thus confirm that the PGE<sub>2</sub> that appears in the ivc immediately after an LPS challenge originates in the liver and is probably due to the postulated activation of C5a by LPS. The secondary elevation of PGE<sub>2</sub> 30–45 min after LPS is presumably due to the Toll-like receptor 4-mediated activation of Kc by LPS and the consequent, delayed upregulation of COX-2/mPGES-1. The essential role of Kc as the cellular source of this PGE<sub>2</sub> had been shown earlier (Blatteis et al., 2004a) and was reconfirmed in this study (Perlik et al., 2005). The association between the uptake by Kc of iv or ip injected LPS and the febrile response and that between the latter and the rise in plasma PGE<sub>2</sub> were also demonstrated in parallel studies in conscious animals (Blatteis et al., 2004a; Li et al., 2006). Especially important in this regard was the finding that PGE<sub>2</sub> antiserum administered iv 10 min before iv LPS prevented the febrile response (Li et al., 2006). In sum, these results strongly support the view that PGE<sub>2</sub> is generated by C5a-activated Kc in immediate response to LPS and that it, rather than pyrogenic cytokines, triggers the febrile response. In view of the rapid onset of fever following the iv administration of LPS and the likelihood reviewed earlier that the pyrogenic signal is conveyed to the VMPO neurally, it is probable that the released PGE<sub>2</sub> acts on hepatic vagal afferents for this transmission, although its binding to these terminals has not yet been specifically demonstrated. Figure 1 summarizes the sequence of events triggered in the

periphery by the iv or ip injection of LPS that initiate the febrile response.

### Contribution of preoptic PGE<sub>2</sub> to fever production

It is well established that the pyrogenic signal of peripheral LPS activates specific neural systems in the VMPO region. The enhanced expression of *c-fos* in the NTS and its blockade by subdiaphragmatic vagotomy were already mentioned. In addition, the central noradrenergic system is activated, causing the release of norepinephrine (NE), an effect that is also blocked by vagotomy (Fleshner et al., 1995; Ishizuka et al., 1997); the increase in the activity of the noradrenergic system produced by proinflammatory stimuli is well documented (for review, see Dunn, 2001). NE microinjected into the VMPO of conscious guinea pigs raises their  $T_c$  (Zeisberger, 1987; Quan and Blatteis, 1989), and electrical stimulation of their ascending noradrenergic system in the NTS produces the same effect (Szelenyi et al., 1976, 1977) whereas chemical sympathectomy prevents this response (Zeisberger, 1987). NE also stimulates the release of PGE<sub>2</sub> in brain tissue *in vitro* (Hori et al., 1987), and the microdialysis of NE into the VMPO of conscious guinea pigs augments the local production of PGE<sub>2</sub> (Sehic et al., 1996a, b; Feleder et al., unpublished observation); in turn, the released PGE<sub>2</sub> inhibits the further presynaptic release of NE (Bergstrom et al., 1973; for review, see Hedqvist, 1977). Hence, NE locally released in the VMPO consequent to the vagally transmitted hepatic PGE<sub>2</sub> signal could induce the production of preoptic PGE<sub>2</sub> and thereby account for its presence and traditional pyrogenic function there. Indeed, the release of NE in the VMPO following the systemic administration of LPS or IL-1 $\beta$  has been demonstrated in several species (Linthorst et al., 1995; Wiczorek et al., 2005; Feleder et al., unpublished observation; for review, see Dunn, 2001). It has further been reported that pretreatment with the non-selective COX inhibitor aspirin prevents the  $T_c$  rise produced by PGE<sub>2</sub> microinjected into a lateral ventricle (Navarro et al., 1988).

If NE were the direct stimulus for the increased production of PGE<sub>2</sub> in the VMPO in response to

peripheral LPS-stimulated, vagally transmitted signals, the induction of PGE<sub>2</sub> should begin promptly and, according to current dogma, be mediated by COX-2/mPGES-1. Indeed, the LPS-induced rise in preoptic PGE<sub>2</sub> occurs promptly following the peripheral injection of LPS (Sehic et al., 1996a, b; Feleder et al., unpublished observation; for review, see Blatteis, 1997). However, the increase in preoptic NE that attends the course of the febrile response to LPS is associated with the early rather than the late phase of fever (Linthorst et al., 1995; Feleder et al., unpublished observation). Hence, the temporal discrepancy between the occurrence of NE and PGE<sub>2</sub> in the VMPO would seem to argue against their functional interaction. There is another challenge: as mentioned earlier, although, in brain, both COX-2 and mPGES-1 are expressed constitutively, but rather minimally, in endothelial cells and, to a lesser extent, in neuronal cell bodies, dendritic spines, and glia, their role in fever production is attributed to their upregulation in endothelial and/or glial cells, but not in neurons (Breder and Saper, 1996; Matsumura et al., 1997; Cao et al., 1998; Quan et al., 1998). Moreover, their mRNAs become detectable in the VMPO between 0.5 and 4 h after the iv or ip administration of LPS or a cytokine (Inoue et al., 2002; Ivanov et al., 2002). The prompt elevation of preoptic PGE<sub>2</sub> could therefore not be due to its COX-2/mPGES-1-mediated production. Consequently, its production should be mediated either via COX-1 or via a COX-independent pathway.

In a series of experiments designed to clarify the mechanism that could underlie the production of PGE<sub>2</sub> in the VMPO in response to local NE and, at the same time, to identify the adrenoceptor (AR) subtype(s) that could be involved in this effect, NE and specific  $\alpha_1$ - and  $\alpha_2$ -AR agonists and antagonists were microdialyzed into the VMPO of conscious guinea pigs pretreated intraVMPO with selective COX-1 and COX-2 inhibitors (Feleder et al., 2004). The results unexpectedly revealed that NE mediates not one, but two successive  $T_c$  rises, each associated with a different mechanism. Thus, the intraVMPO microdialysis of the selective  $\alpha_1$ -AR agonist cirazoline induced a prompt  $T_c$  rise without, remarkably, affecting basal preoptic PGE<sub>2</sub> levels, whereas the selective  $\alpha_2$ -AR agonist

clonidine caused a significantly delayed  $T_c$  rise that followed an initial  $T_c$  fall; both these responses were accompanied by parallel changes in the levels of VMPO PGE<sub>2</sub>. The elevation of PGE<sub>2</sub>, however, was not associated under these conditions with a demonstrable upregulation of COX-2 (Feleder et al., unpublished observation), implying, therefore, that it was mediated by the activation of this enzyme, not its de novo synthesis. The thermal effects of both agonists were validated by their blockade by their respective, selective antagonists, prazosin and yohimbine. Furthermore, both the increases in  $T_c$  and preoptic PGE<sub>2</sub> levels caused by clonidine were prevented by the intraVMPO microdialysis of a selective COX-2 inhibitor, MK-0663; they were unaffected by that of the selective COX-1 inhibitor SC-560, although it unexpectedly suppressed the initial decreases of both these variables caused by this  $\alpha_2$ -AR agonist. The intraVMPO microdialysis of NE reproduced the early and the late  $T_c$  and preoptic PGE<sub>2</sub> level changes induced by clonidine, but not the early  $T_c$  rise evoked by cirazoline; both the clonidine-mediated effects were inhibited by yohimbine and MK-0663, but not by prazosin. Cirazoline and clonidine microdialyzed together replicated the late  $T_c$  rises but not the early  $T_c$  falls elicited by clonidine and NE, and also not the early  $T_c$  rises caused by cirazoline; on the other hand, their co-microdialysis induced both the VMPO PGE<sub>2</sub> falls and rises associated with clonidine and NE. In further support of the non-involvement of PGE<sub>2</sub> in the cirazoline-induced  $T_c$  rise and, in contrast, the involvement of COX-2-dependent PGE<sub>2</sub> in that caused by clonidine, conscious, wild-type, and COX-1<sup>-/-</sup> mice respond similarly to the intracerebroventricular (icv) microinjection of these two  $\alpha$ -AR agonists as the correspondingly treated guinea pigs; and, most relevant to this context, the late clonidine-induced  $T_c$  rise is absent in COX-2<sup>-/-</sup> mice (Blatteis et al., 2004a).

The finding that the specific  $\alpha_1$ -AR agonist cirazoline evoked very quickly a rise in  $T_c$  without the intermediation of PGE<sub>2</sub> was indeed remarkable and unexpected. It suggested that the  $\alpha_1$ -AR activated by NE are located on postsynaptic warm-sensitive or thermoinsensitive neurons in the VMPO and that NE directly reduces or augments,

respectively, the activities of these neurons; according to the classical model of Hammel (1965); both responses promote heat conservation. Since these neurons, moreover, are thought to inhibit synaptically connected cold-sensitive neurons, these are concomitantly facilitated, stimulating heat production; i.e., in combination, these effector mechanisms raise  $T_c$ . Support for the direct, PGE<sub>2</sub>-independent involvement of  $\alpha_1$ -AR is also provided by preliminary data from single-unit extracellular recordings of the discharge rates of thermally characterized, individual neurons in horizontal slices of rat hypothalami. In the experiments to date, cirazoline inhibited three of four warm-sensitive neurons and excited two of three thermoinsensitive neurons (Boulant and Blatteis, unpublished observation). The specific  $\alpha_1$ -AR subtype involved in this hyperthermic effect remains to be identified.

The findings that, when the microdialysis of clonidine was continued for another 3 h, the initial hypothermic response was followed by a protracted  $T_c$  rise and an associated increase in the animals' levels of preoptic PGE<sub>2</sub> were both also novel. Since both these responses were also inhibited by yohimbine pretreatment, they were therefore also  $\alpha_2$ -AR-mediated; and since, moreover, they were prevented by the prior microdialysis of the selective COX-2 inhibitor MK-0663, this late hyperthermic response was specifically mediated by COX-2-dependent PGE<sub>2</sub>. The brain cell type expressing COX-2 in response to NE in this context remains to be determined. But, since the increase of COX-2 following the peripheral administration of LPS has been observed, as already mentioned, in glial and cerebrovascular endothelial cells, but only irregularly in neurons, we conjecture that the PGE<sub>2</sub> collected in the microdialysate effluents from the VMPO interstitial space in these experiments was generated by astrocytic processes contacting noradrenergic synaptic regions rather than by postsynaptic (warm-sensitive) neurons. The subsequent effect of the thus released PGE<sub>2</sub> on the electrical activities of VMPO warm-sensitive and thermoinsensitive neurons is presumptively similar to that indicated earlier for  $\alpha_1$ -AR-mediated responses, viz., a reduction and an increase in their discharge rates, respectively. The PGE<sub>2</sub>-sensitive receptor involved in these neuronal effects is probably the EP<sub>3</sub> subtype;

it has been linked to the development of fever and is present in the VMPO (Ushikubi et al., 1998; Ek et al., 2000; for review, see Oka, 2004). The receptor implicated in the inhibition of presynaptic NE release by PGE<sub>2</sub> has also been previously identified as the EP<sub>3</sub> subtype (Schlicker and Gothert, 1998). These results thus demonstrated that the induction of PGE<sub>2</sub> by NE in the VMPO is modulated by  $\alpha_2$ -AR and indeed catalyzed by COX-2. The identity of the  $\alpha_2$ -AR subtype involved in this effect remains to be elucidated.

Taken together, the preceding data would suggest, therefore, that the NE released in the VMPO in response to the vagally conveyed pyrogenic message of Kc-generated PGE<sub>2</sub> could mediate the febrile response of guinea pigs in the following two, successive ways: (1) it could induce the first of the characteristic two  $T_c$  rises evoked in conscious guinea pigs by iv LPS by rapidly activating  $\alpha_1$ -AR without the intermediation of PGE<sub>2</sub>, and (2) it could cause the second of the two  $T_c$  rises and also the delayed, single rise produced by ip LPS (due to the late arrival of LPS in the liver [Li and Blatteis, 2004]) by also stimulating at the same time  $\alpha_2$ -AR, consequently activating (after the delay imposed by the de novo synthesis of the enzymes) the production and release of COX-2/mPGES-1-dependent PGE<sub>2</sub> in the POA. This hypothesis was tested (Feleder et al., unpublished observation) by measuring over short intervals the febrile response and the levels of NE and PGE<sub>2</sub> in the interstitial fluid of the VMPO of conscious guinea pigs pretreated with intraVMPO prazosin or yohimbine (or their vehicles) to the iv or Ip injection of *Salmonella enteritidis* LPS, following the same protocol as in the preceding study. In guinea pigs, pyrogenic doses of iv injected LPS rapidly and characteristically evoke two successive  $T_c$  rises associated with concurrent elevations in preoptic PGE<sub>2</sub> levels (Sehic et al., 1996a, b); however, the second rise is attenuated significantly more than the first by COX-2 blockade (Steiner et al., 2001). Ip injected LPS, on the other hand, induces coincident monophasic  $T_c$  and PGE<sub>2</sub> responses after some delay (Inoue et al., 2002). The iv injection of a low-to-moderate dose of LPS (2  $\mu$ g/kg) promptly induced the appearance of NE in the VMPO interstitial fluid; its level culminated in 30 min, then gradually

returned toward its control value over the following 2 h.  $T_c$  and preoptic PGE<sub>2</sub> levels also both increased promptly, first rapidly, then more slowly, and in correspondence with each other; the first  $T_c$  peak was reached in ~60 min and the second in ~150 min. Pretreatment with the  $\alpha_1$ -AR antagonist prazosin significantly slowed the rate of rise of the early phase of fever and eliminated the first  $T_c$  peak; but it did not affect the onset latency of the febrile response nor the magnitude and time of the second peak of fever. It also did not affect the initial, LPS-induced elevation of preoptic PGE<sub>2</sub>; but, importantly, this increase was not sustained: PGE<sub>2</sub> levels returned from their first highs at 30 min to their control values at 60 min. They then resumed their rise, culminating not differently than their untreated counterparts and coincidentally with the second peak of fever (Feleder et al., unpublished observations). These results thus confirmed that the onset of fever evoked in conscious guinea pigs by iv LPS is associated with the intraVMPO release of NE and accompanied by coincident increases in  $T_c$  and preoptic PGE<sub>2</sub> levels, in conformity with previous findings (Linthorst et al., 1995; Sehic et al., 1996a, b; Wieczorek et al., 2005). They further demonstrated that the initial  $T_c$  rise is indeed mediated by the NE-induced activation of  $\alpha_1$ -AR and intimated that, its very onset apparently excepted, the first phase of fever was maintained by the direct noradrenergic activation of the relevant neurons in the VMPO, without the intermediation of PGE<sub>2</sub>. Pretreatment with the  $\alpha_2$ -AR antagonist yohimbine, by contrast, did not change the onset and rate of rise of the early febrile response, the first  $T_c$  peak reaching the same value and occurring at the same time as that of the untreated controls; but it completely abrogated the second peak, consequently significantly reducing the overall magnitude of the febrile response by comparison with that of untreated controls. The fever, moreover, abated more slowly in this group than in its controls. Remarkably, this treatment completely suppressed the LPS-induced rise in preoptic PGE<sub>2</sub> levels (Feleder et al., unpublished observations). These findings thus indicated that the LPS fever of these  $\alpha_2$ -AR-blocked guinea pigs is initiated, maintained, and even extended in the total absence of

corresponding increases in VMPO PGE<sub>2</sub> levels. Hence, by deduction, it is mediated entirely by PGE<sub>2</sub>-independent,  $\alpha_1$ -AR activation. Neither the microdialysis of prazosin nor of yohimbine *per se* affected the animals'  $T_c$  and preoptic PGE<sub>2</sub> levels (Feleder et al., unpublished observations). In sum, it would appear that the early phase of LPS fever is indeed mediated independently of PGE<sub>2</sub> by  $\alpha_1$ -AR stimulation whereas the late phase is dependent on PGE<sub>2</sub> induced consequent to  $\alpha_2$ -AR stimulation, as hypothesized. But then what accounts for the initial rise in preoptic PGE<sub>2</sub> of all the animals, including the yohimbine-pretreated ones?

The first possibility is that COX-1 could mediate the initial, LPS-induced PGE<sub>2</sub> rise. But this possibility was refuted by findings that the intraVMPO microdialysis of the selective COX-1 inhibitor SC-560 had absolutely no effect on the onset, height, and course of  $T_c$  and the associated changes in preoptic PGE<sub>2</sub> levels induced in conscious guinea pigs by iv LPS (Feleder et al., unpublished observation). Acetaminophen, a putatively selective inhibitor of COX-1 variant retaining intron 1 (for review, see Botting and Ayoub, 2005), also did not affect the  $T_c$  and intraVMPO PGE<sub>2</sub> responses to iv LPS (Feleder et al., unpublished observation). Administration of the selective COX-2 inhibitor MK-0663, by contrast, did not alter the early  $T_c$  and preoptic PGE<sub>2</sub> rises, but prevented both their late rises (Feleder et al., unpublished observation). These results, therefore, documented again that the late, but not the early phase of LPS fever is mediated by COX-2-dependent PGE<sub>2</sub>, and that COX-1 evidently plays no role in the febrile response (except, presumptively in part, in the initial induction of PGE<sub>2</sub> by C5a-stimulated Kc, as mentioned earlier).

Since the intraVMPO microdialysis of selective COX-1, -2, and -3 inhibitors do not prevent the prompt, initial increase in PGE<sub>2</sub> levels observed in the POA/VMPO following the peripheral administration of LPS, it may be surmised that this rise is induced by a COX-independent pathway, i.e., presumptively by the non-enzymatic isoprostane pathway of free radical-catalyzed peroxidation of AA (for review, see Basu and Helmersson, 2005). Indirect support for the involvement of free radicals in the observed, initial PGE<sub>2</sub> elevation comes from the finding that the intraVMPO

microdialysis of the antioxidant catechin throughout the febrile course had the same effect as pretreatment with the  $\alpha_2$ -AR antagonist yohimbine, i.e., suppression of both the LPS-induced early and late rises of preoptic PGE<sub>2</sub> and of the second peak of fever; the latency of fever onset and its first peak were not affected, the fever continuing at its early phase high level until it abated normally (Feleder et al., unpublished observation). The free radicals in this case could be generated by the auto-oxidation of NE and/or nitric oxide (NO). The latter is also released locally in the VMPO after LPS administration (see below); however, no direct measurement of biomarkers of oxidative stress was made in these studies. It is not clear at this writing whether catechin also inhibited the expression of COX-2 and, hence, the production of COX-2-dependent PGE<sub>2</sub> (Wang et al., 2004). But it has been reported previously that fever is prevented when the production of free radicals is blocked. The effect was ascribed then not to the inhibition by antioxidants of isoprostane production, but rather to their reduction of the thiol groups attached to *N*-methyl-D-aspartate receptors, thereby depressing glutamate-mediated neuronal excitability and, hence, limiting fever (Riedel, 1997; Riedel and Maulik, 1999; Riedel et al., 2003); the activation by peripheral pyrogens of glutaminergic pathways both in the NTS and the OVLT has been reported (Mascarucci et al., 1998; Huang et al., 2001, 2004). It was therefore suggested that NO and oxygen radicals, but not

PGE<sub>2</sub>, modulate fever (Riedel et al., 2003). Indeed, since the initial  $T_c$  rise provoked by the peripheral administration of LPS is evidently mediated by PGE<sub>2</sub>-independent  $\alpha_1$ -AR activation in the VMPO and since, moreover, the febrile rise can evidently be sustained by this mechanism alone until its normal abatement, it raises the heretic possibility that, in contrast to PGE<sub>2</sub> generated in the liver, PGE<sub>2</sub> in the VMPO, however it may occur there, may not be material to the febrile response!

Finally, reports that the gaseous transmitter NO stimulates the biosynthesis of PGE<sub>2</sub> by increasing the activities of both isoforms of COX prompted us to investigate whether NO could also have a pyretic function in the central mediation of the febrile response. Indeed, the various isoforms of NO synthase (endothelial, neural, and inducible NOS), the enzyme that converts L-arginine into citrulline and NO, occur in the hypothalamus (for review, see Mollace et al., 2005), and circulating LPS and cytokines stimulate the release of NO in the VMPO (for review, see Schmid et al., 1998; Simon, 1998; Gerstberger, 1999). Furthermore, as reviewed above, NE induces the production of COX-2/mPGES-1-dependent PGE<sub>2</sub> in the VMPO via an  $\alpha_2$ -AR-mediated mechanism, and others have shown that NE activates NOS (Canteros et al., 1996). Hence, the existence of a pyrogenic NE-NO-PGE<sub>2</sub> cascade in the POA in response to iv LPS seems plausible. But the testing of this hypothesis revealed that, quite to the contrary, NO donors microdialyzed into the POA of conscious

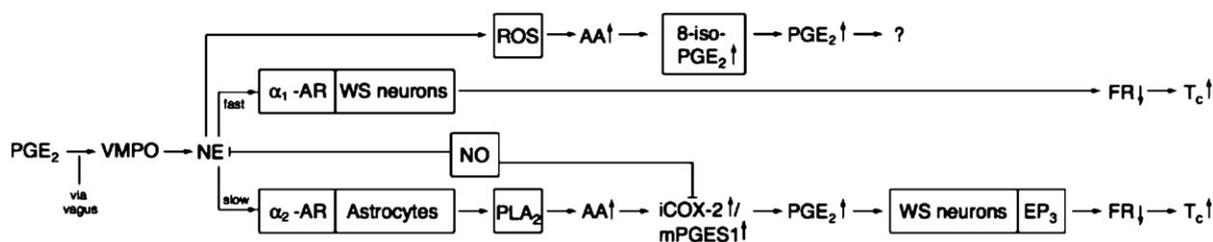


Fig. 2. Schematic illustration of the dual mechanisms, one fast,  $\alpha_1$ -AR-mediated and PGE<sub>2</sub>-independent, the other slow,  $\alpha_2$ -AR-mediated and COX-2/mPGES-1-derived PGE<sub>2</sub>-dependent, activated by NE liberated in the VMPO. Both mechanisms sequentially lead to the depression of the firing rates (FR) of local warm-sensitive (WS) neurons, thereby causing  $T_c$  to rise. NO inhibits the release of NE and may also inhibit the upregulation of COX-2. Reactive oxygen species (ROS) due to the auto-oxidation of NE or derived from NO may also induce isoprostanes (8-iso-PGE<sub>2</sub>) that are quickly converted to PGE<sub>2</sub>; it, however, does not appear to contribute to the initial  $T_c$  rise. Circulating pyrogenic cytokines and cerebrovascular endothelial cells-generated PGE<sub>2</sub> probably contribute to sustaining the course of the late phase of fever.

guinea pigs inhibit rather than promote the febrile response to iv LPS and that they do so by inhibiting the LPS-induced release of NE in the VMPO and consequently preventing the  $\alpha_2$ -AR-mediated activation of COX-2-dependent PGE<sub>2</sub> synthesis (Feleder et al., unpublished observation). NO scavengers microdialyzed into the POA have exactly the opposite effects (Feleder et al., unpublished observation). Indeed, previous data regarding a potential role of NO in fever have been conflicting, some indicating pyretic and others antipyretic effects (Riedel, 1997; Gerstberger, 1999; Kozak and Kozak, 2003). The present results would indicate, therefore, that NO, presumably released in the VMPO coincidentally with or very shortly after NE, serves as a local negative-regulatory factor, i.e., it is a central, endogenous antipyretic mediator of LPS fever. Figure 2 illustrates the sequence of events occurring in the VMPO following the arrival of the peripheral PGE<sub>2</sub>-induced, vagally transmitted, pyrogenic signal.

## Conclusions

LPS-induced fever arises as the result of a complex, phased sequence of interactions among soluble factors and cells that is initiated in the periphery and then transmitted neurally to the VMPO, which modulates the febrile response. Thus, novel evidence accumulated over the past 10 years suggests that the febrigenic process is initiated by the arrival of LPS in the liver and its uptake by Kc, causing virtually immediately the activation of the C cascade and, hence, the generation of the anaphylatoxin C5a. It, in turn, stimulates the Kc very rapidly to release constitutive COX-1- and COX-2-dependent PGE<sub>2</sub>. The released PGE<sub>2</sub> activates local sensory vagal terminals that project to the NTS. From the NTS, the input of PGE<sub>2</sub> is transmitted to the VMPO via the ventral noradrenergic bundle. NE consequently secreted in the VMPO activates both local  $\alpha_1$ - and  $\alpha_2$ -AR. The stimulation of the first rapidly evokes an initial rise in  $T_c$  which is associated with a decrease in the firing rates of preoptic warm-sensitive neurons, inhibiting heat loss and stimulating heat

production, but is not accompanied by any change in the levels of preoptic PGE<sub>2</sub>. Stimulation of the second causes, after a significant delay, a second, more prolonged  $T_c$  rise that is associated with a concurrent increase in preoptic COX-2/mPGES-1-dependent PGE<sub>2</sub> levels. Hence, two distinctly produced PGE<sub>2</sub>s would appear to mediate the febrile response: one, generated in the liver, is the immediate distal trigger of the febrile response, and the other, produced in the VMPO, is its subsequent proximal modulator. The late phase is probably also supported by meanwhile produced circulating cytokines and PGE<sub>2</sub> as well as PGE<sub>2</sub> generated by cerebral endothelial and/or other cells, all acting presumably as originally proposed for their roles.

In summary, the key, new findings regarding the initiation of LPS fever are: (1) peripheral PGE<sub>2</sub> rather than pyrogenic cytokines initiates the febrile response, (2) NE propagates the pyrogenic signal forward within the VMPO, (3) NO modulates its release in the VMPO and, hence, the intensity of the febrile response, and (4) COX-2/mPGES-1-dependent PGE<sub>2</sub> mediates the late, but not the early phase of fever; the latter appears to be independent of COX-derived PGE<sub>2</sub>.

## Acknowledgments

This research was supported by National Institute of Health grants Nos. NS-34857 and NS-38594. I gratefully acknowledge the invaluable contributions of my co-workers in this research, Drs. Elmir Sehic, Shuxin Li, Carlos Feleder, Vit Perlik, Zhonghua Li, and Ying Tang, without whose hard work and unswerving dedication these studies could not have been performed.

## References

- Abul, H.T., Davidson, J., Milton, A.S. and Rotondo, D. (1997) Prostaglandin E<sub>2</sub> enters the brain following stimulation of the acute phase immune response. *Ann. N.Y. Acad. Sci.*, 813: 287–295.
- Ambs, P., Baccarini, M., Fitzke, E. and Dieter, P. (1995) Role of cytosolic phospholipase A2 in arachidonic acid release of rat-liver macrophages: regulation by Ca<sup>2+</sup> and phosphorylation. *Biochem. J.*, 311: 189–195.

- Basu, S. and Helmersson, J. (2005) Factors regulating isoprostane formation in vivo. *Antioxid. Redox Signal*, 7: 221–235.
- Bergstrom, S., Farnebo, L.O. and Fuxe, K. (1973) Effect of prostaglandin E<sub>2</sub> on central and peripheral catecholamine neurons. *Eur. J. Pharmacol.*, 21: 362–368.
- Blatteis, C.M. (1997) Prostaglandin E<sub>2</sub>: a putative fever mediator. In: Mackowiak P.A. (Ed.), *Fever: Basic Mechanisms and Management* (2nd ed.). Lippincott-Raven, Philadelphia, PA, pp. 117–145.
- Blatteis, C.M., Li, S., Li, Z., Perlik, V. and Feleder, C. (2004a) Complement is required for the induction of endotoxic fever in guinea pigs and mice. *J. Thermal Biol.*, 9: 369–381.
- Blatteis, C.M., Li, S., Li, Z., Perlik, V. and Feleder, C. (2004b) Signaling the brain in systemic inflammation: the role of complement. *Front. Biosci.*, 9: 915–931.
- Blatteis, C.M. and Sehic, E. (1997) Fever: how may circulating pyrogens signal the brain? *News Physiol. Sci.*, 12: 1–7.
- Botting, R. and Ayoub, S.A. (2005) COX-3 and the mechanism of action of paracetamol/acetaminophen. *Prostaglandins Leukot. Essent. Fatty Acids*, 72: 85–87.
- Boulant, J.A. (2000) Role of the preoptic-anterior hypothalamus in thermoregulation and fever. *Clin. Infect. Dis.*, 31(Suppl 5): S157–S161.
- Breder, C.D. and Saper, C.B. (1996) Expression of inducible cyclooxygenase mRNA in the mouse brain after systemic administration of bacterial lipopolysaccharide. *Brain Res.*, 713: 64–69.
- Canteros, G., Rettori, V., Genaro, A., Suburo, A., Gimeno, M. and McCann, S.M. (1996) Nitric oxide synthase content of hypothalamic explants; increased by norepinephrine and inactivated by NO and cGMP. *Proc. Natl. Acad. Sci. U.S.A.*, 93: 4246–4250.
- Cao, C., Matsumura, K., Yamagata, K. and Watanabe, Y. (1998) Cyclooxygenase-2 is induced in brain blood vessels during fever evoked by peripheral or central administration of tumor necrosis factor. *Brain Res. Mol. Brain Res.*, 56: 45–56.
- Conti, B., Tabarean, I., Andrei, C. and Bartfai, T. (2004) Cytokines and fever. *Front. Biosci.*, 9: 1433–1449.
- Dinarello, C.A. (2004) Infection, fever, and exogenous and endogenous pyrogens: some concepts have changed. *J. Endotoxin Res.*, 10: 201–222.
- Dunn, A.J. (2001) Effects of cytokines and infections on brain neurochemistry. In: Ader R., Felten D.L. and Cohen N. (Eds.), *Psychoneuroimmunology*. Academic Press, New York, pp. 645–666.
- Dunn, A.J. (2002) Mechanisms by which cytokines signal the brain. *Int. Rev. Neurobiol.*, 52: 43–65.
- Ek, M., Arias, C., Sawchenko, P. and Ericsson-Dahlstrand, A. (2000) Distribution of the EP3 prostaglandin E<sub>2</sub> receptor subtype in the rat brain: relationship to sites of interleukin-1-induced cellular responsiveness. *J. Comp. Neurol.*, 428: 5–20.
- Ek, M., Kurosowa, M., Lundeberg, T. and Ericsson, A. (1998) Activation of vagal afferents after intravenous injection of interleukin-1 $\beta$ : role of endogenous prostaglandins. *J. Neurosci.*, 18: 9471–9479.
- Feleder, C., Li, Z., Perlik, V., Evans, A. and Blatteis, C.M. (2003) The spleen modulates the febrile response of guinea pigs to LPS. *Am. J. Physiol.*, 284: R1466–R1476.
- Feleder, C., Perlik, V. and Blatteis, C.M. (2004) Preoptic  $\alpha_1$ - and  $\alpha_2$ -noradrenergic agonists induce, respectively, PGE<sub>2</sub>-independent and PGE<sub>2</sub>-dependent hyperthermic responses in guinea pigs. *Am. J. Physiol.*, 286: R1156–R1166.
- Fleshner, M., Goehler, L.E., Hermann, J., Relton, J.K., Maier, S.F. and Watkins, L.R. (1995) Interleukin-1 $\beta$ -induced corticosterone elevation and hypothalamic NE depletion is vagally mediated. *Brain Res. Bull.*, 37: 605–610.
- Fleshner, M., Goehler, L.E., Schwartz, B.A., McGorry, M., Martin, D., Maier, S.F. and Watkins, L.R. (1998) Thermogenic and corticosterone responses to intravenous cytokines (IL-1 $\beta$  and TNF- $\alpha$ ) are attenuated by subdiaphragmatic vagotomy. *J. Neuroimmunol.*, 86: 134–141.
- Gaykema, R.P.A., Goehler, L.E., Tilders, F.J.H., Bol, J.G.J.M., McGorry, M., Fleshner, M., Maier, S.F. and Watkins, L.R. (1998) Bacterial endotoxin induces Fos immunoreactivity in primary afferent neurons of the vagus nerve. *Neuroimmunomodulation*, 5: 234–240.
- Gerstberger, R. (1999) Nitric oxide and body temperature control. *News Physiol. Sci.*, 14: 30–36.
- Goehler, L.E., Relton, J.K., Dripps, D., Kiechle, R., Tartaglia, N., Maier, S.F. and Watkins, L.R. (1997) Vagal paranglia bind biotinylated interleukin-1 receptor antagonists (IL-1ra) in the rat: a possible mechanism of immune-to-brain communication. *Brain Res. Bull.*, 43: 357–364.
- Goldbach, J.M., Roth, J. and Zeisberger, E. (1997) Fever suppression by subdiaphragmatic vagotomy in guinea pigs depends on the route of pyrogen administration. *Am. J. Physiol.*, 272: R675–R681.
- Hammel, H.T. (1965) Neurons and temperature regulation. In: Yamamoto W.S. and Brobeck J.R. (Eds.), *Physiological Controls and Regulations*. W.B. Saunders, Philadelphia, PA, pp. 71–97.
- Hansen, M.K., O'Connor, K.A., Goehler, L.E., Watkins, L.R. and Maier, S.F. (2001) The contribution of the vagus nerve in interleukin-1beta-induced fever is dependent on dose. *Am. J. Physiol.*, 280: R929–R934.
- Hedqvist, P. (1977) Basic mechanisms of prostaglandin action on autonomic neurotransmission. *Annu. Rev. Pharmacol. Toxicol.*, 17: 259–279.
- Hori, Y., Blatteis, C.M. and Nasjletti, A. (1987) Production of PGE<sub>2</sub> by brain slices stimulated by various thermoactive agents. *Fed. Proc.*, 46: 683.
- Huang, W.T., Tsai, S.M. and Lin, M.T. (2001) Involvement of brain glutamate in pyrogenic fever. *Neuropharmacology*, 41: 811–818.
- Huang, W.T., Wang, J.J. and Lin, M.T. (2004) Cyclooxygenase inhibitors attenuate augmented glutamate release in organum vasculosum laminae terminalis and fever induced by staphylococcal enterotoxin A. *J. Pharmacol. Sci.*, 94: 192–196.
- Inoue, W., Matsumura, K., Yamagata, K., Takemiya, T., Shiraki, T. and Kobayashi, S. (2002) Brain-specific endothelial induction of prostaglandin E<sub>2</sub> synthesis enzymes: its temporal relation to fever. *Neurosci. Res.*, 44: 51–61.

- Ishizuka, Y., Ishida, Y., Kunitake, T., Kato, K., Hanamori, T., Mitsuyama, Y. and Kannan, H. (1997) Effects of area postrema lesion and abdominal vagotomy on interleukin-1 $\beta$ -induced norepinephrine release in the hypothalamic paraventricular nucleus region in the rat. *Neurosci. Lett.*, 223: 57–60.
- Ivanov, A.I., Pero, R.S., Scheck, A.C. and Romanovsky, A.A. (2002) Prostaglandin E (2)-synthesizing enzymes in fever: differential transcriptional regulation. *Am. J. Physiol.*, 283: R1104–R1117.
- Ivanov, A.I. and Romanovsky, A.A. (2004) Prostaglandin E<sub>2</sub> as a mediator of fever: synthesis and catabolism. *Front. Biosci.*, 9: 1977–1993.
- Jansky, L., Vybiral, S., Pospisilova, D., Roth, J., Dornand, J. and Zeisberger, E. (1995) Production of systemic and hypothalamic cytokines during the early phase of endotoxin fever. *Neuroendocrinology*, 62: 55–61.
- Kozak, W. and Kozak, A. (2003) Genetic models in applied physiology: selected contributions: differential role of nitric oxide synthase isoforms in fever of different etiologies: studies using Nos gene-deficient mice. *J. Appl. Physiol.*, 94: 2534–2544.
- Li, Z. and Blatteis, C.M. (2004) Fever onset is linked to the appearance of lipopolysaccharide in the liver. *J. Endotoxin Res.*, 10: 1–15.
- Li, Z., Feleder, C. and Blatteis, C.M. (2004) Lipopolysaccharide challenge causes exaggerated fever and increased hepatic lipopolysaccharide uptake in vinblastine-induced leukopenic guinea pigs. *Crit. Care Med.*, 32: 2131–2134.
- Li, Z., Perlik, V., Feleder, C., Tang, Y. and Blatteis, C.M. (2006) Kupffer cell-generated PGE<sub>2</sub> triggers the febrile response of guinea pigs to intravenously injected LPS. *Am. J. Physiol.*, 290: R1262–R1270.
- Linthorst, A.C., Flachskamm, C., Holsboer, F. and Reul, J.M. (1995) Intraperitoneal administration of bacterial endotoxin enhances noradrenergic neurotransmission in the rat preoptic area: relationship with body temperature and hypothalamic–pituitary–adrenocortical axis activity. *Eur. J. Neurosci.*, 7: 2418–2430.
- Mascarucci, P., Perego, C., Terrazzino, S. and De Simoni, M.G. (1998) Glutamate release in the nucleus tractus solitarius induced by peripheral lipopolysaccharide and interleukin-1 $\beta$ . *Neuroscience*, 86: 1285–1290.
- Matsumura, K., Cao, C. and Watanabe, Y. (1997) Possible role of cyclooxygenase-2 in the brain vasculature in febrile response. *Ann. N.Y. Acad. Sci.*, 813: 302–306.
- Mollace, V., Muscoli, C., Masini, E., Cuzzocrea, S. and Salve-mini, D. (2005) Modulation of prostaglandin biosynthesis by nitric oxide and nitric oxide donors. *Pharmacol. Rev.*, 57: 217–252.
- Morimoto, A., Morimoto, K., Watanabe, T., Sakata, Y. and Murakami, N. (1992) Does an increase in prostaglandin E<sub>2</sub> in the blood circulation contribute to a febrile response in rabbits? *Brain Res. Bull.*, 29: 189–192.
- Navarro, E., Romero, S.D. and Yaksh, T.L. (1988) Release of prostaglandin E<sub>2</sub> from brain of cat: II. In vivo studies on the effects of adrenergic, cholinergic and dopaminergic agonists and antagonists. *Neuropharmacology*, 27: 1067–1072.
- Nijima, A. (1996) The afferent discharges from sensors for interleukin-1 $\beta$  in the hepatoportal system in the anesthetized rat. *J. Auton. Nerv. Syst.*, 61: 287–291.
- Oka, T. (2004) Prostaglandin E<sub>2</sub> as a mediator of fever: the role of prostaglandin E (EP) receptors. *Front. Biosci.*, 9: 3046–3057.
- Perlik, V., Li, Z., Goorha, S., Ballou, L.R. and Blatteis, C.M. (2005) LPS-activated complement, not LPS per se, triggers the early release of PGE<sub>2</sub> by Kupffer cells. *Am. J. Physiol.*, 289: R332–R339.
- Quan, N. and Blatteis, C.M. (1989) Intraoperatively microdialyzed and microinjected norepinephrine evokes different thermal responses. *Am. J. Physiol.*, 257: R816–R821.
- Quan, N., Whiteside, M. and Herkenham, M. (1998) Cyclooxygenase-2 mRNA expression in rat brain after peripheral injection of lipopolysaccharide. *Brain Res.*, 802: 189–197.
- Rhur, S.G. (1994) Regulation of phosphoinositide-specific phospholipase C by G protein. In: Liscovitch M. (Ed.), *Signal-Activated Phospholipases*. RG Landes, Austin, TX, pp. 1–12.
- Riedel, W. (1997) Antipyretic role of nitric oxide during endotoxin-induced fever in rabbits. *Int. J. Tissue React.*, 19: 171–178.
- Riedel, W., Lang, U., Oetjen, U., Schlapp, U. and Shibata, M. (2003) Inhibition of oxygen radical formation by methylene blue, aspirin, or  $\alpha$ -lipoic acid, prevents bacterial-lipopolysaccharide-induced fever. *Mol. Cell Biochem.*, 247: 83–94.
- Riedel, W. and Maulik, G. (1999) Fever: an integrated response of the central nervous system to oxidative stress. *Mol. Cell Biochem.*, 196: 125–132.
- Romanovsky, A.A., Ivanov, A.I. and Karman, E.K. (1999) Blood-borne, albumin-bound prostaglandin E<sub>2</sub> may be involved in fever. *Am. J. Physiol.*, 276: R1840–R1844.
- Romanovsky, A.A., Kulchitsky, V.A., Simons, C.T., Sugimoto, N. and Szekely, M. (1997b) Febrile responsiveness of vagotomized rats is suppressed even in the absence of malnutrition. *Am. J. Physiol.*, 273: R777–R783.
- Romanovsky, A.A., Simons, C.T., Szekely, M. and Kulchitsky, V.A. (1997a) The vagus nerve in the thermoregulatory response to systemic inflammation. *Am. J. Physiol.*, 273: R407–R413.
- Roth, J. and De Souza, G.E. (2001) Fever induction pathways: evidence from responses to systemic or local cytokine formation. *Braz. J. Med. Biol. Res.*, 34: 301–314.
- Rotondo, D., Abul, H.T., Milton, A.S. and Davidson, J. (1988) Pyrogenic immunomodulators increase the level of prostaglandin E<sub>2</sub> in the blood simultaneously with the onset of fever. *Eur. J. Pharmacol.*, 154: 145–152.
- Saper, C.B. (1998) Neurobiological basis of fever. *Ann. N.Y. Acad. Sci.*, 856: 90–94.
- Schieferdecker, H.L., Rothermel, E., Timmermann, A., Gotze, O. and Jungermann, K. (1997) Anaphylatoxin C5a receptor mRNA is strongly expressed in Kupffer and stellate cells and weakly in sinusoidal endothelial cells but not in hepatocytes of normal rat liver. *FEBS Lett.*, 406: 305–308.
- Schieferdecker, H.L., Schlaf, G., Jungermann, K. and Gotze, O. (2001) Functions of anaphylatoxin C5a in rat liver: direct

- and indirect actions on nonparenchymal and parenchymal cells. *Int. Immunopharmacol.*, 1: 469–481.
- Schlaf, G., Schmitz, M., Rothermel, E., Jungermann, K., Schieferdecker, H.L. and Gotze, O. (2003) Expression and induction of anaphylatoxin C5a receptors in the rat liver. *Histol. Histopathol.*, 18: 299–308.
- Schlicker, E. and Gothert, M. (1998) Interactions between the presynaptic alpha2-autoreceptor and presynaptic inhibitory heteroreceptors on noradrenergic neurones. *Brain Res. Bull.*, 47: 129–132.
- Schmid, H.A., Riedel, W. and Simon, E. (1998) Role of nitric acid in temperature regulation. *Prog. Brain Res.*, 115: 25–49.
- Schütze, S. and Krönki, M. (1994) Activation of phosphatidylcholine-specific phospholipase C by cytokines. In: Liscovitch M. (Ed.), *Signal-Activated Phospholipases*. RG Landes, Austin, TX, pp. 101–123.
- Sehic, E. and Blatteis, C.M. (1996) Blockade of lipopolysaccharide-induced fever by subdiaphragmatic vagotomy in guinea pigs. *Brain Res.*, 726: 160–166.
- Sehic, E., Szekely, M., Ungar, A.L., Oladehin, A. and Blatteis, C.M. (1996a) Hypothalamic PGE<sub>2</sub> during lipopolysaccharide-induced fever in guinea pigs. *Brain Res. Bull.*, 39: 391–399.
- Sehic, E., Ungar, A.L. and Blatteis, C.M. (1996b) Interaction between norepinephrine and prostaglandin E<sub>2</sub> in the preoptic area of guinea pigs. *Am. J. Physiol.*, 271: R528–R536.
- Simon, E. (1998) Nitric oxide as a peripheral and central mediator in temperature regulation. *Amino Acids*, 14: 87–93.
- Simons, C.T., Kulchitsky, V.A., Sugimoto, N., Homer, L.D., Szekely, M. and Romanovsky, A.A. (1998) Signaling the brain in systemic inflammation: which vagal branch is involved in fever genesis? *Am. J. Physiol.*, 275: R63–R68.
- Skarnes, R.C., Brown, S.K., Hull, S.S. and McCracken, J.A. (1981) Role of prostaglandin E in the biphasic fever response to endotoxin. *J. Exp. Med.*, 154: 1212–1224.
- Steiner, A.A., Li, S., Llanos-Q, J. and Blatteis, C.M. (2001) Differential inhibition by nimesulide of the early and late phases of intravenous- and intracerebroventricular-LPS-induced fever in guinea pigs. *Neuroimmunomodulation*, 9: 263–275.
- Szelenyi, Z., Zeisberger, E. and Bruck, K. (1976) Effects of electrical stimulation in the lower brainstem on temperature regulation in the unanaesthetized guinea-pig. *Pflugers Arch.*, 364: 123–127.
- Szelenyi, Z., Zeisberger, E. and Bruck, K. (1977) A hypothalamic alpha-adrenergic mechanism mediating the thermogenic response to electrical stimulation of the lower brainstem in the guinea pig. *Pflugers Arch.*, 370: 19–23.
- Ushikubi, F., Segi, E., Sugimoto, Y., Murata, T., Matsuoka, T., Kobayashi, T., Hizaki, H., Tuboi, K., Katsuyama, M., Ishikawa, A., Tanaka, T., Yoshida, N. and Narumiya, S. (1998) Impaired febrile response in mice lacking the prostaglandin E receptor subtype EP3. *Nature*, 395: 281–284.
- Wan, W., Wetmore, L., Sorensen, C.M., Greenberg, A.H. and Nance, D.M. (1994) Neural and biochemical mediators of endotoxin and stress-induced *c-fos* expression in the rat brain. *Brain Res. Bull.*, 34: 7–14.
- Wang, T., Qin, L., Liu, B., Liu, Y., Wilson, B., Eling, T.E., Langenbach, R., Taniura, S. and Hong, J.-S. (2004) Role of reactive oxygen species in LPS-induced production of prostaglandin E<sub>2</sub> in microglia. *J. Neurochem.*, 88: 939–947.
- Watkins, L.R., Goehler, L.E., Reston, J.K., Tartaglia, N., Gilbert, L., Martin, D. and Maier, S.F. (1995) Blockade of interleukin-1-induced hyperthermia by subdiaphragmatic vagotomy: evidence for vagal mediation of immune-brain communication. *Neurosci. Lett.*, 183: 27–31.
- Wieczorek, M., Swiergiel, A.H., Pournajafi-Nazarloo, H. and Dunn, A.J. (2005) Physiological and behavioral responses to interleukin-1 $\beta$  and LPS in vagotomized mice. *Physiol. Behav.*, 85: 500–511.
- Zeisberger, E. (1987) The roles of monoaminergic neurotransmitters in thermoregulation. *Can. J. Physiol. Pharmacol.*, 65: 1395–1401.